Corneal Scraping for Acanthamoeba

Arrangements

Follow the special instruction listed below, if you have any questions please contact the F.C. Blodi Eye Pathology Laboratory (319-335-7095) prior to beginning this procedure.

- Make sure the following materials are in the room prior to performing the scraping:
  - **i.** If you are using the **Saccomanno or cytology Fixative**
    - **a)** Spatula for scraping the cornea
    - **b)** Sterile container (centrifuge tube, specimen container)
    - **c)** Wash bottle containing Saccomanno fixative
    - **d)** Ocular Pathology Consultation Request form with the proper basic patient information, clinical data, and specimen location (ie: corneal scrapping right eye) written on the form.
  - **ii.** If you are using the **10% Neutral buffered formalin method:**
    - **a)** Spatula for scraping the cornea
    - **b)** Slides with one end frosted
    - **c)** Coplin jar with 10% neutral buffered formalin
    - **d)** Paper toweling
- The following procedural instructions based on the materials available to your institution are:
  - **i.** If you have **Saccomanno or cytology fixative** available use these instructions (this is the preferred method):
    - **a)** Have an open sterile container (centrifuge tube, specimen container) available prior to scraping the patient.
    - **b)** Using a spatula scrape the patient as usual.
    - **c)** Immediately after scraping the patient, using wash bottle containing fixative, wash cells and tissue off spatula and into the sterile container.
    - **d)** Make sure not to use more than 0.5 ml of fixative to wash the cells off the spatula.
    - **e)** Past experience has shown when more than 0.5 ml of fixative is used, the lab produces 12 cytospin slides. There is usually a sparse cell count per slide. When using 0.5 ml Saccomanno, the lab produces 6 slides with a better cell representation per slide.
    - **f)** Take tip of spatula and swirl in the Saccomanno to assure that all of the cells have been removed from the spatula.
    - **g)** Place the lid on the container tightly
    - **h)** Label the specimen container with the patient's name and date of birth
  - **ii.** If you are using **10% Neutral Buffered Formalin**:
    - **a)** Scrape the cornea, immediately smear the scrapings on the clear portion of a clean glass slide. The specimen should be smeared so that the frosted end is facing up.
    - **b)** Immediately immerse the slide containing the specimen in the coplin jar containing 10% Neutral Buffered Formalin.
    - **c)** Allow the slides to sit in the 10% Neutral Buffered Formalin fixative for 15-20 minutes.

Note: **DO NOT** leave the slides in the formalin more than 15-20 minutes or the tissue may fall off the slides.

- ❖ Remove the slides from the fixative.
- ❖ Place the slides on a paper towel to air dry.

Note: Make sure the side of the slide containing the specimen is right side up or the scraping may be wiped off.

- ❖ After **completely** air dried, place the slides into a slide mailer (can be provided by the F.C. Blodi Eye Pathology laboratory).
- ❖ Fill out the Ocular Pathology Consultation Request form **completely**.